

# Using SmartTag as operational welfare indicator of farmed fish

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*Using SmartTag as operational welfare indicator of  
farmed fish*

# 1) What is SmartTag?

# What is SmartTag?



The SmartTag provides online measurements of breathing pattern in free-swimming fish

- Monitor and document fish welfare status in aquaculture
- Early warning system
- Optimize production regimes
- Assess feed intake rates



# What is SmartTag? (II)

1.



## 1) SmartTag prototype

- 46 x 16 mm, 6 g in water / 15 g in air
- 60 - 130 kHz acoustic transmission
- produced by THELMA, Norway

2a



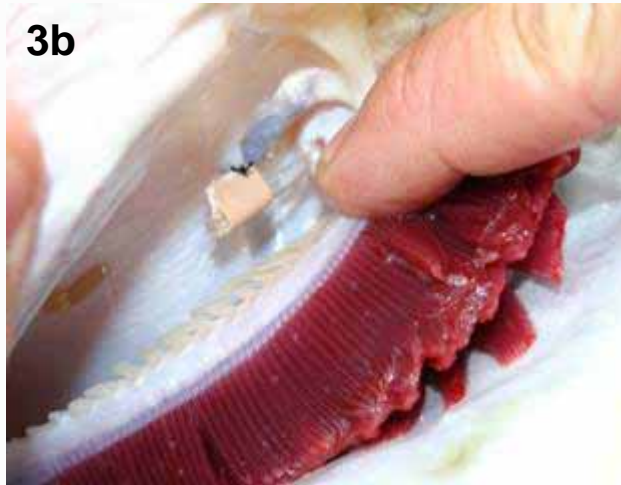
2ab) The tag is attached externally, on the back of the anaesthetized fish

2b





# What is SmartTag? (II)

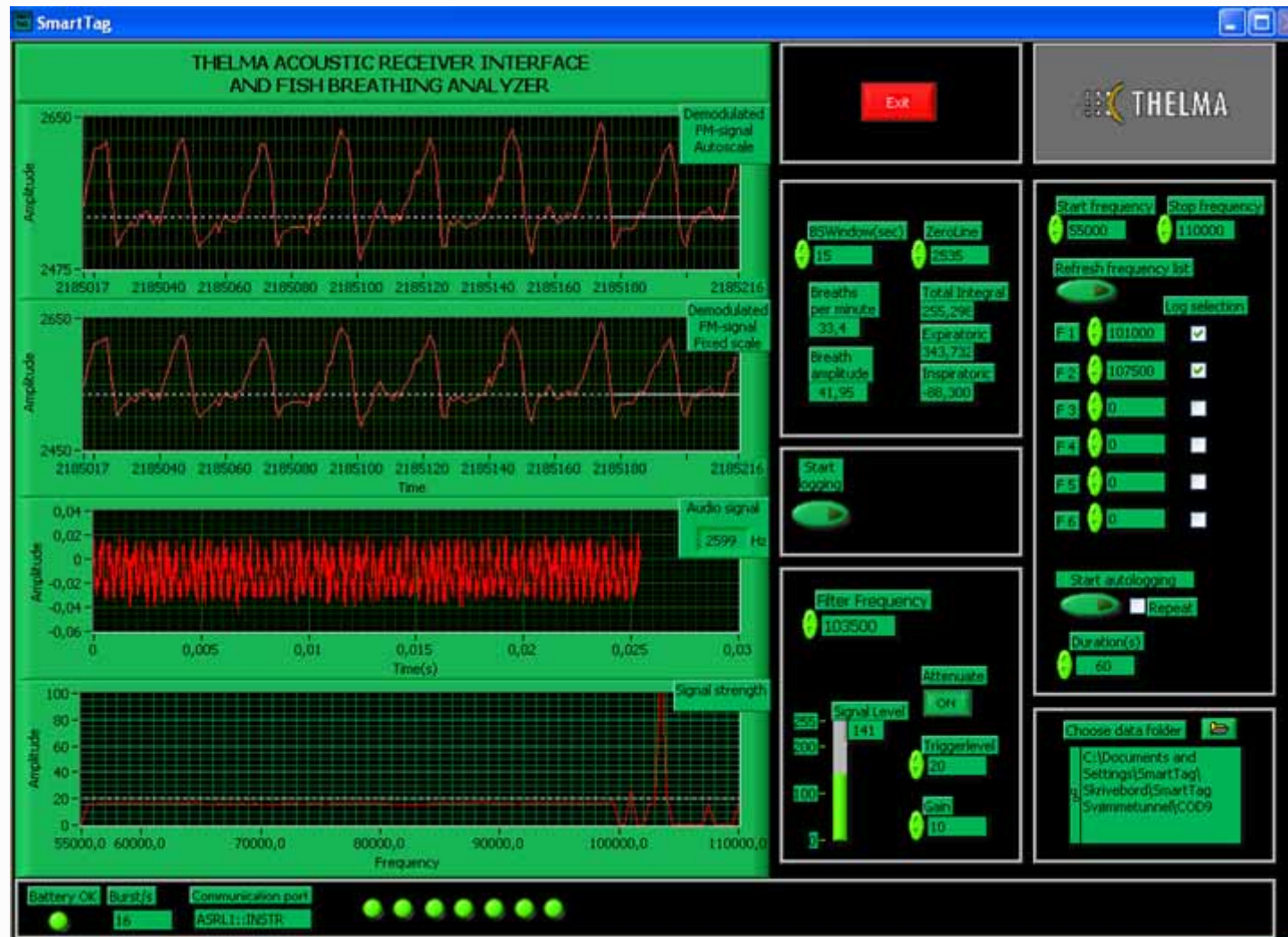


3abc)

A water-filled tube (TYGON 1.6 mm ID) is sutured inside the mouth and connected to the tag for online pressure measurements, thus producing detailed data on fish breathing activity.



# What is SmartTag? (III)



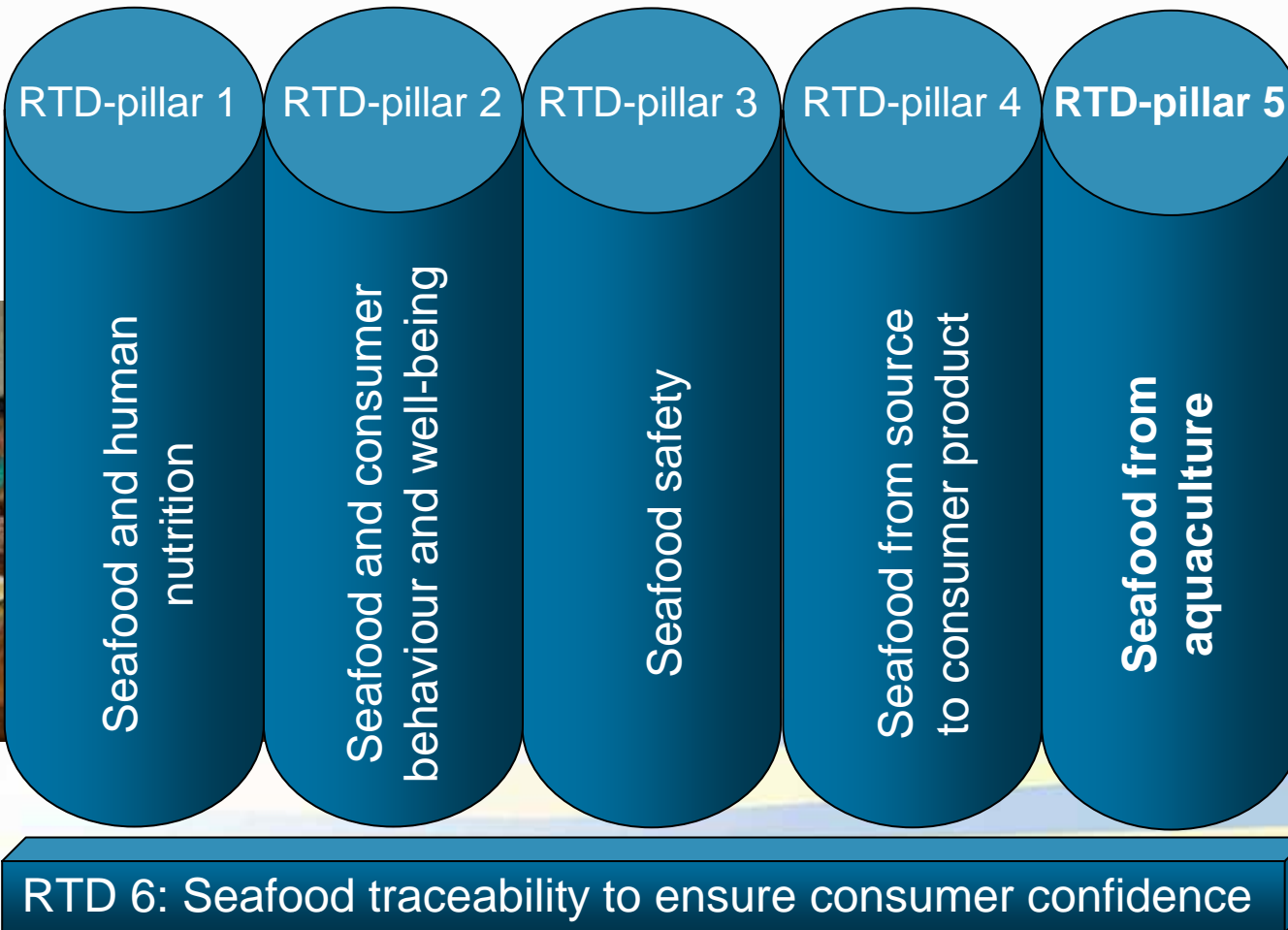
*The fish breathing is visualised and stored on a computer*

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farmed fish*

## **2) Background**

# SEAFOODplus:

A value-chain - “fork to fish farm” perspective





# Trends in today's aquaculture (I)

- Larger fish production units
- More intensive production:
  - more fish pr volume water
  - artificial oxygenation, accumulation of CO<sub>2</sub> and other metabolites
- More off-shore and submerged production facilities
- Rationalization of man-work  
(fewer people pr kg fish produced)



Large off-shore facility



Submerged facility

# Trends in today's aquaculture (II)



- Increased scepticism among consumers on food safety and animal welfare issues (Damsgård 2006)
- *Need for monitoring and documenting fish welfare and rearing conditions*



Fish farm



Product offered consumers /  
retailers

# Monitoring and documenting fish welfare in aquaculture



## Traditional approach (1):

- Veterinarian inspections and analyzes of sick or dead fish



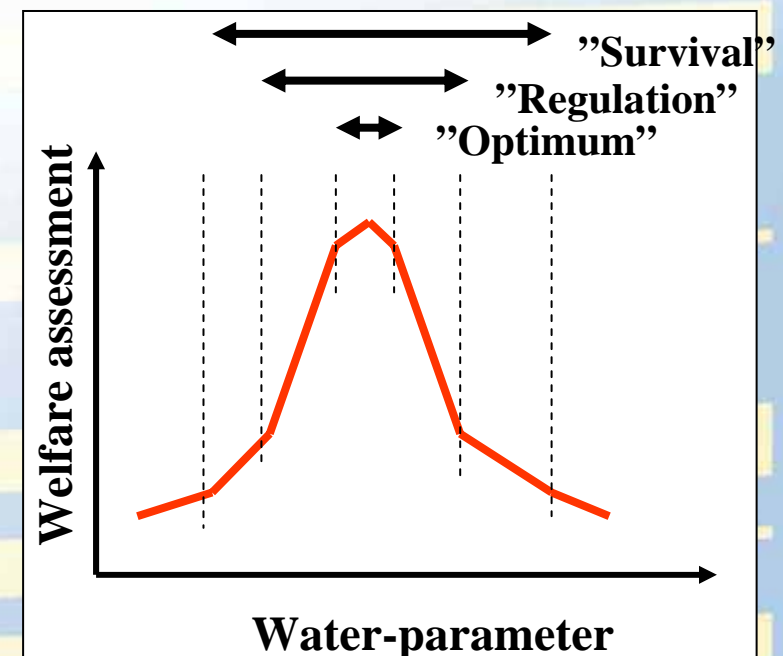
- *Periodical, and/or based on prior suspicion of something being wrong, and may often be too late*

# Monitoring and documenting fish welfare in aquaculture



## Traditional approach (2):

- Monitor environmental factors and ensure that these are within a pre-determined, acceptable range
- *Does not take in to account the “sum of factors”, and that fish responses to such factors are variable*



***New approach:***

## **SmartTag and the “canary cod” principle (I)**



Canary birds were used as indicators of air quality in mines

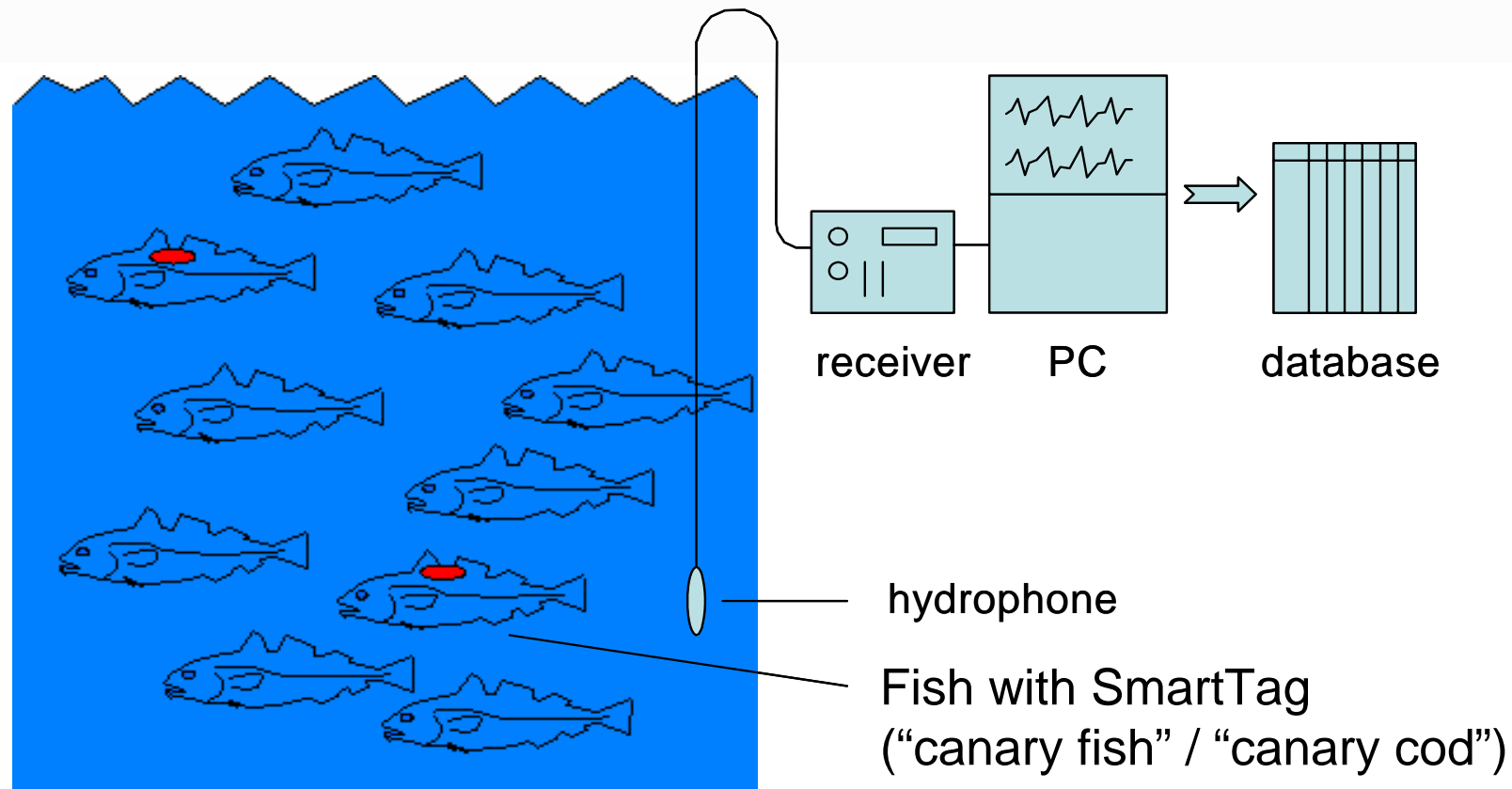
*Small cage with canary bird used in testing for carbon monoxide gas in Hollinger Mine, Timmons, Ontario, Canada (<http://www.msha.gov>)*



# SmartTag and the “canary cod” principle (II)



- Online monitoring of how individual fish respond to their aquaculture environment



*A physiological parameter is continuously monitored by the tags, picked up by a reception system, and processed online on a computer with relevant parameters being stored automatically in a database.*

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### **3) Validation of the SmartTag system and of fish breathing as welfare indicator**

# Validation of the SmartTag system:

## A. Existing scientific literature



*We know from the scientific literature that fish breathing activity responds to factors such as:*

- Hypoxia, hypercapnea and water pH (e.g. Smith & Jones 1982; Reid et al. 2000)
- Toxic or sub-toxic levels of metabolites and xenobiotics in feed and water (e.g. Pane et al. 2004)
- Parasite infection, disease, anaemia (e.g. Byrne et al. 1991)
- General stress response (e.g. Laitinen et al. 1996)
- Fear and pain (e.g. Sneddon et al. 2003)

# Validation of the SmartTag system:

## B. Controlled experiments

### I) Responses to water quality manipulation

- SmartTagged Atlantic cod (*Gadus morhua*) swimming in a swim tunnel respirometer
- Breathing activity at normal (excellent) water quality is compared with breathing activity during short-term exposure to different water qualities relevant to the aquaculture industry

### II) Responses to handling disturbance (pilot studies)

- Free-swimming, SmartTagged Atlantic cod are exposed to a handling disturbance
- Fish breathing activity compared before, during and after stress

# 1. Responses to water quality manipulation: experimental design

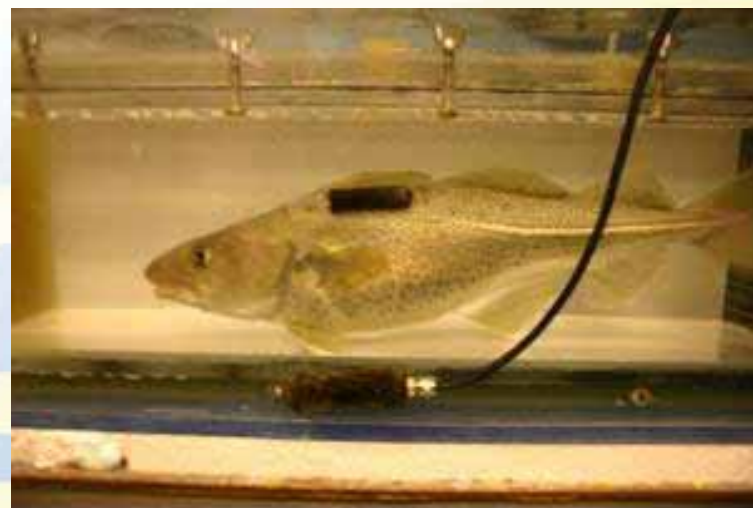


## Experimental day 1:

Fish SmartTagged and introduced to swim tunnel respirometer at normal (excellent) water quality



Experimental room with swim tunnel



SmartTagged 1.5 kg cod swimming at  $0.5 \text{ BLs}^{-1}$  at  $10^\circ\text{C}$  in the swim tunnel respirometer.



# 1. Responses to water quality manipulation: experimental design (II)



## Experimental day 2:

- 1) Control measurements (3 x 0.5 hours)
- 2) Treatment exposure (A, B, C, D, E or F) at low level (3 x 0.5 hours)
- 3) Treatment exposure (as above) at high level (3 x 0.5 hours)
- 4) Blood sampling

## Treatments (water qualities):

- |                        |   |
|------------------------|---|
| A. Control             | D. High CO <sub>2</sub>                               |
| B. Low O <sub>2</sub>  | E. Low O <sub>2</sub> + high CO <sub>2</sub> (B + D)  |
| C. High O <sub>2</sub> | F. High O <sub>2</sub> + high CO <sub>2</sub> (C + D) |

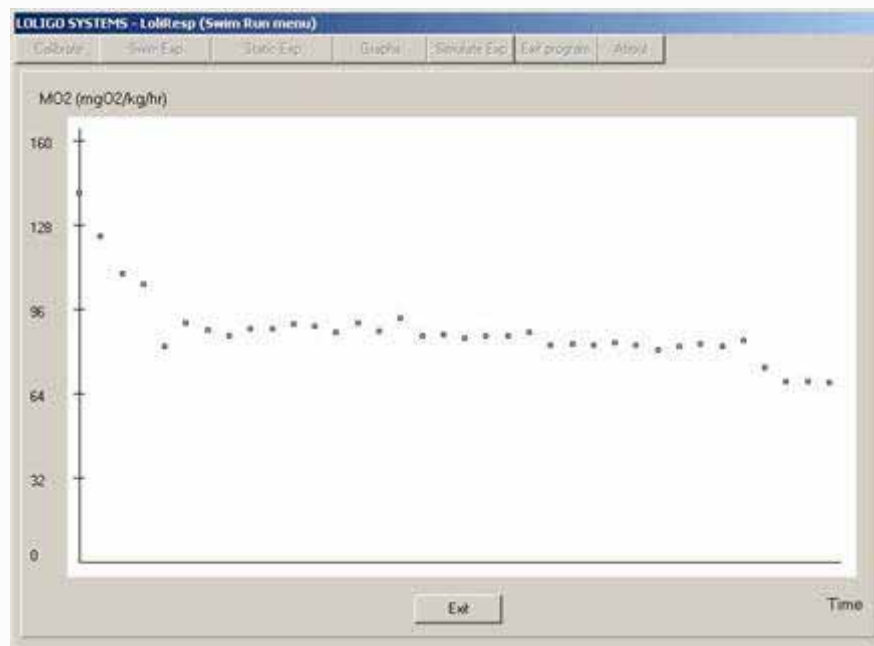
**Blood** is sampled for cortisol (stress hormone) analysis at day 1, and for analyses of cortisol, glucose, lactate, ions and gas tensions at day 2.

# 1. Responses to water quality manipulation: experimental design (III)

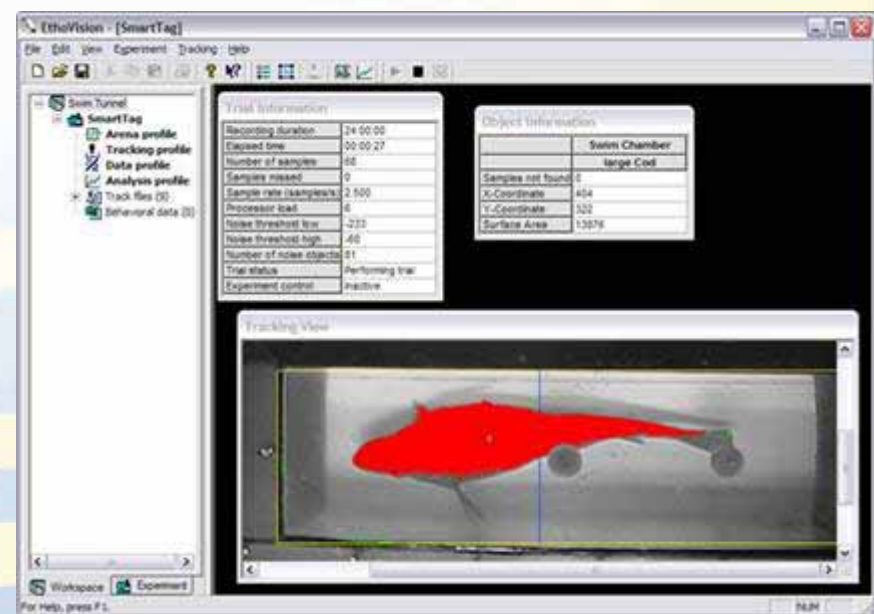


## Experimental day 1 *and* 2:

- Continuous registration of fish metabolism (30 min cycles)
- Continuous registration of fish behaviour (online video analysis)



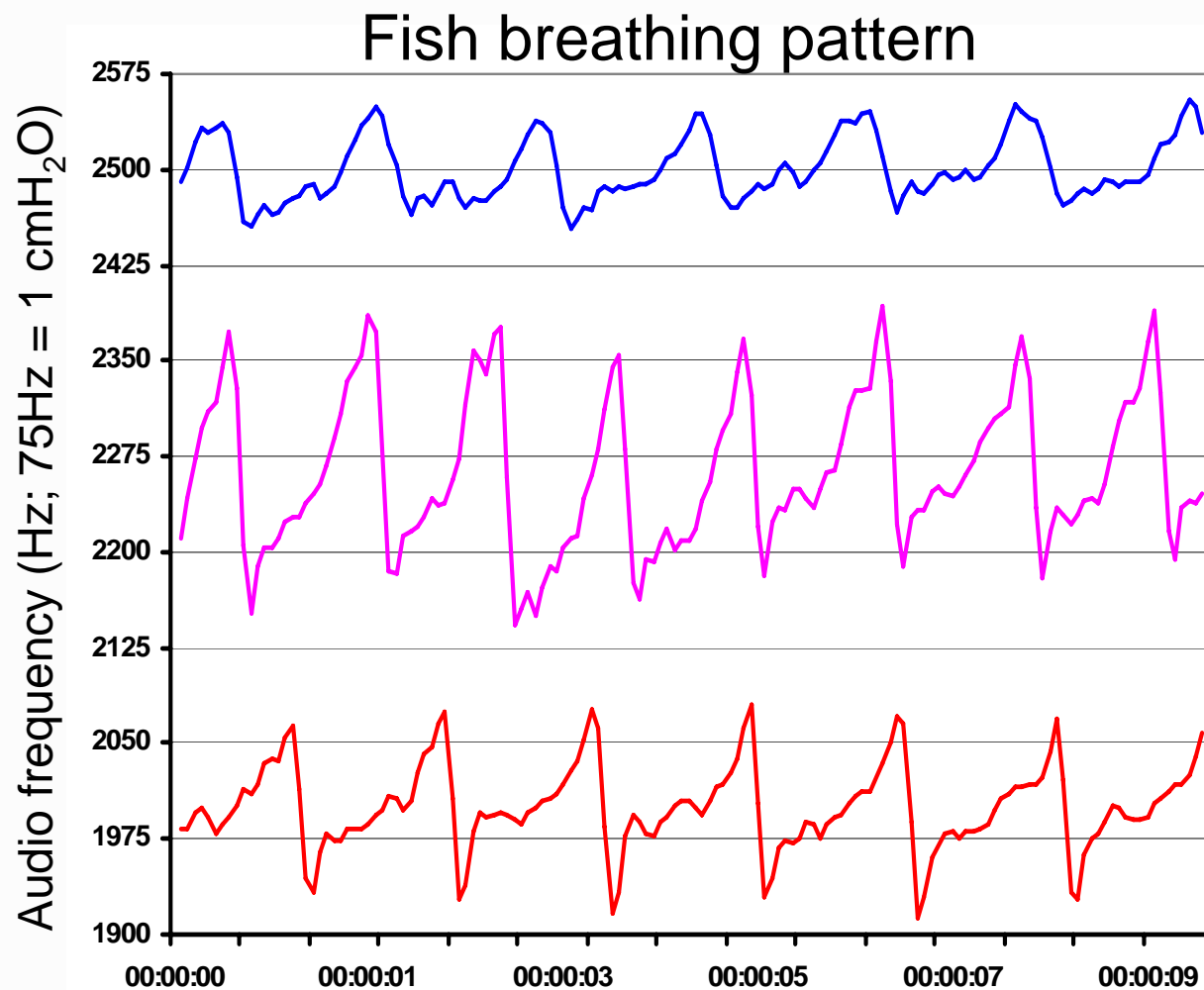
Changes in metabolism following introduction into the swim tunnel respirometer



Automatic video-tracking and analysis of fish behaviour in the swim tunnel respirometer

# 1. Responses to water quality manipulation:

Low O<sub>2</sub> levels (e.g. sea cage culture)



**Normoxia; O<sub>2</sub> = 100 %**

- Frequency = 38 min<sup>-1</sup>
- Amplitude = 1.2 cm H<sub>2</sub>O

**Hypoxia; O<sub>2</sub> = 65%**

- Frequency = 47 min<sup>-1</sup>
- Amplitude = 3.0 cm H<sub>2</sub>O

**Hypoxia; O<sub>2</sub> = 45%**

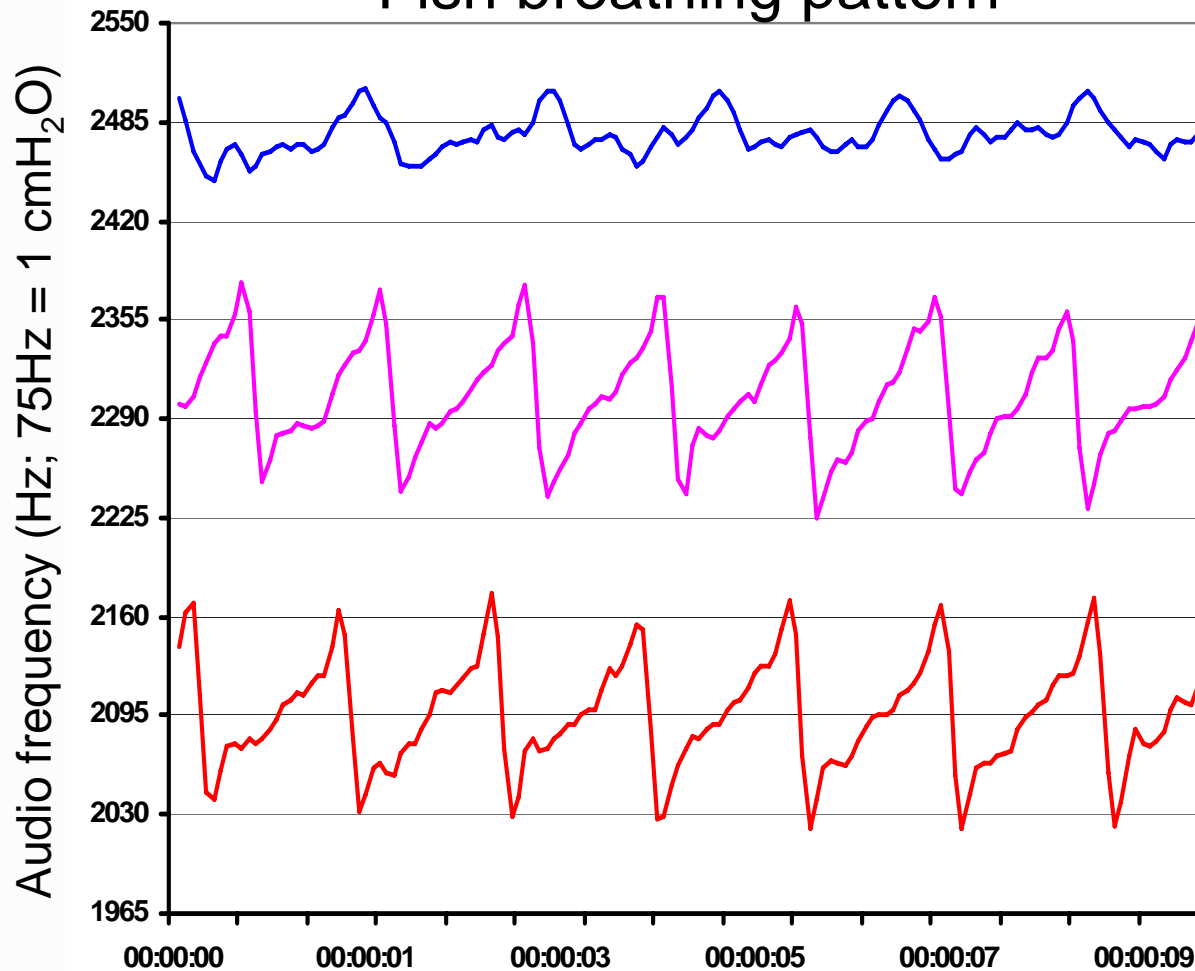
- Frequency = 41
- Amplitude = 2.3 cmH<sub>2</sub>O

# 1. Responses to water quality manipulation:

High CO<sub>2</sub> levels (e.g. recirculation systems)



Fish breathing pattern



## Normal water

(CO<sub>2</sub> = 1 mg/L)

- Frequency = 35 min<sup>-1</sup>
- Amplitude = 0.8 cm H<sub>2</sub>O

## CO<sub>2</sub> = 17 mg/L

- Frequency = 45 min<sup>-1</sup>
- Amplitude = 2.2 cm H<sub>2</sub>O

## CO<sub>2</sub> = 30 mg/L

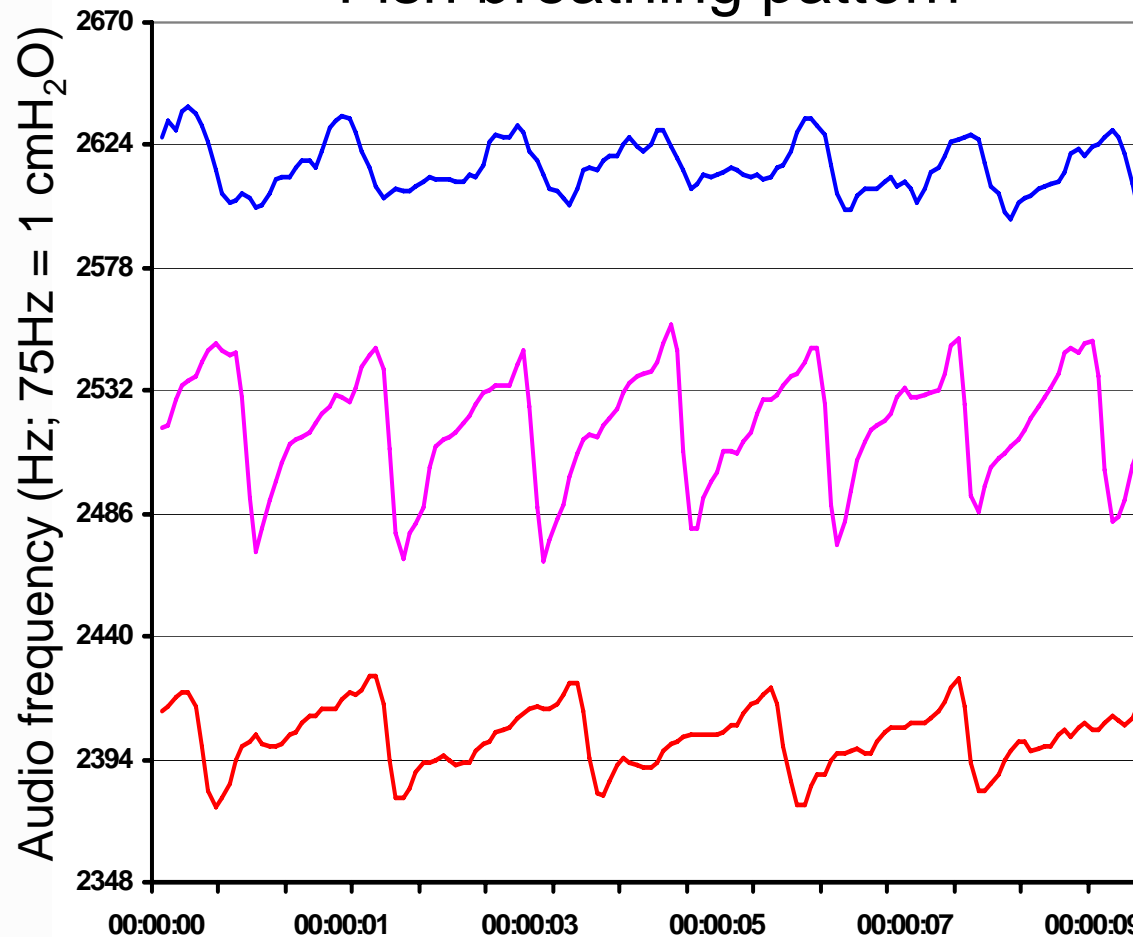
- Frequency = 41 min<sup>-1</sup>
- Amplitude = 2.4 cmH<sub>2</sub>O

# 1. Responses to water quality manipulation:

low O<sub>2</sub> levels + high CO<sub>2</sub> (e.g. pond culture)



## Fish breathing pattern



### Normal water

(O<sub>2</sub>= 100%, CO<sub>2</sub>= 1mg/L)

- Frequency= 38 min<sup>-1</sup>
- Amplitude = 0.8 cm H<sub>2</sub>O

### O<sub>2</sub>= 65% + CO<sub>2</sub>= 13mg/L

- Frequency= 42 min<sup>-1</sup>
- Amplitude = 1.7 cm H<sub>2</sub>O

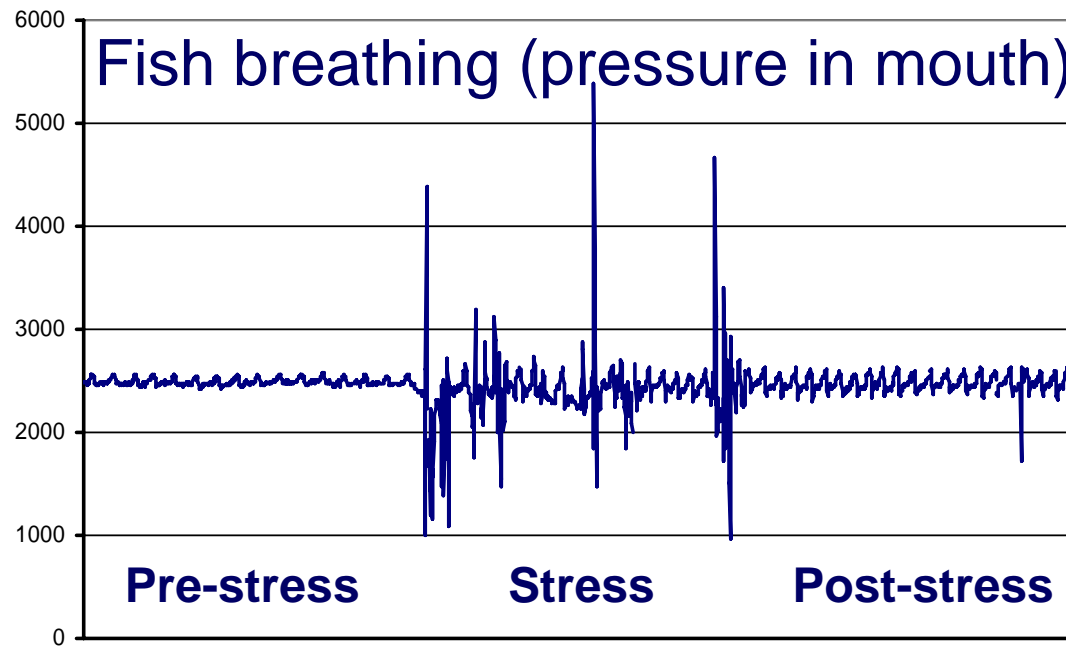
### O<sub>2</sub>= 45% + CO<sub>2</sub>= 35 mg/L

- Frequency= 31 min<sup>-1</sup>
- Amplitude = 1.1 cm H<sub>2</sub>O

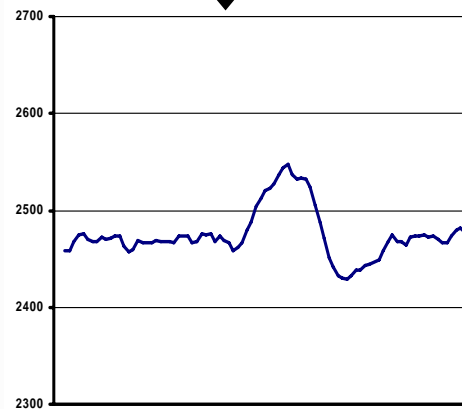


# Validation of SmartTag:

## 2. Responses to handling disturbance



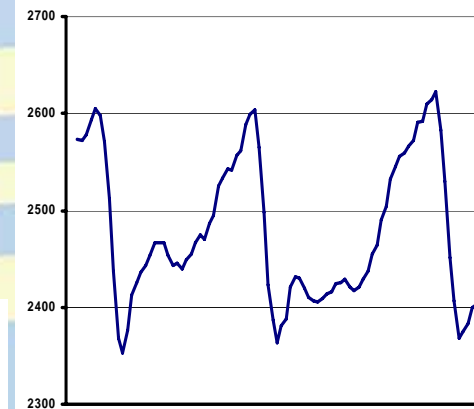
Method: 2 min dip-netting in small sea cage



**Stress:**  
RF = ?,  $\Delta P$  up to 50 cmH<sub>2</sub>O

RF = 16  
 $\Delta P$  = 1 cmH<sub>2</sub>O

RF = 22  
 $\Delta P$  = 3 cmH<sub>2</sub>O



# Using SmartTag as operational welfare indicator of farmed fish - CONCLUSIONS



- The SmartTag system is a promising candidate operational welfare indicator of farmed fish
- May also function as an early warning system and be used to optimize production regimes
- Further development and validation is needed before full-scale usage of the SmartTag system



# Acknowledgements



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# A better life with seafood...



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